Expression Profile of Apoptotic Genes Bak1 and Bcl2 in Pakistani Presbycusis Patients from Faisalabad and Chichawatni Populations

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ABSTRACT

OBJECTIVE: To analyze the expression profiles of two apoptotic genes, BAK1 and BCL2, in Pakistani ARHI patients' blood samples compared to healthy subjects.

METHODOLOGY: This expression profiles study was conducted at the Sight Centre of Bahawalpur City of Southern Punjab from August 2022 to July 2023. All of the subjects were from Faisalabad or Chichawatni, Pakistan cities. The participants exposed to head injury or trauma, noise exposure, ototoxic medication, chemical exposure, diabetes, liver cirrhosis, neurological disease, psychiatric disease, cognitive dysfunction, middle ear disease, and brain tumour factors were excluded from the study. Individuals with cold or flu were also excluded from the study. RNA was extracted from both the peripheral blood samples of healthy and affected individuals. The quantity of RNA was then determined, and cDNA was synthesized from it. This cDNA was then utilized in qPCR to assess the relative expression of the BAK1. BCL2, and ACTB genes using gene-specific primers.

RESULTS: Our results suggested that ARHI patients had higher levels of BAK1 expression and BAK1/ BCL2 ratio than healthy subjects.

CONCLUSION: Thus, apoptosis mediated by BAK1 may be a key mechanism controlling the development of ARHI. Furthermore, changes in BAK1 gene expression of blood samples might be utilized as a rapid test for early diagnosis of ARHI.

KEYWORDS: Presbycusis, Age related hearing impairment (ARHI), Apoptosis, BAK1, BCL2

INTRODUCTION

Age-related hearing impairment (ARHI), commonly known as presbycusis, is a more prevalent multifactorial disorder among aged people. With an increase in the expectancy of human life and the ratio of elderly persons, the incidence of ARHI is also elevating¹. Individuals aged 65 years or above have this hearing impairment. It is considered an untreatable and unpreventable disorder². ARHI is a crucial health concern with a considerable social impact. Age-related hearing impairment (ARHI) is related to cognition, which has grabbed the attention of many researchers who have studied this disease³. This disease has been linked with anxiety, dementia and depression. It is a bilateral, symmetrical, and sensorineural higher frequency impairment in which the hearing threshold tends to damage slowly with age. Gradual degeneration of various sensory cells

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increases with age and poor diet⁴. This disease is caused by damage and complete loss of stria vascularis cells, spiral ganglion cells, and sensory hair cells. Some critical mechanisms involved in this hearing loss are oxidative stress, apoptosis, mitochondrial dysfunction and inflammation. The primary reason for ARHI is the loss or impairment of spiral ganglion cells and hair cells of the inner ear⁵. ARHI is now widely acknowledged to be associated with ageing-related reductions in the auditory system's capacity for spectral and temporal resolution. Agerelated variables that decrease neuronal synchronization are thought to be linked to impaired temporal processing⁶.

Apoptosis involves extrinsic and intrinsic signalling pathways. The BCL2 protein family regulates the intrinsic pathway. This family's genes are categorized into two groups: anti-apoptotic and pro-apoptotic. BCL2 and BCLXL are anti-apoptotic genes, while BAK1, BAX, and BAD are pro-apoptotic genes⁷.

BAK1 and Casp3 are apoptosis-related genes whose upregulation has been observed in mice models during the onset of ARHI⁸. The BAK1 gene accelerates the permeability of the outer membrane of mitochondria, creating big pores that enable cytochrome c release and consequently activate the caspase cascade. Cell death induced by the BAK1 gene is considered a key mechanism in the development of ARHI disease. Variations in BAK1 expression can be exploited as a biomarker for the

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early diagnosis of ARHI in peripheral blood samples. Cell destruction of hair cells in the ganglion neurons and cochlea was reduced after the deletion of the *BAK1* gene, delaying the onset of ARHI⁹. Critical factors in the ageing of the inner ear in mice are identified as high expression of mRNA of *BCLX* and *BCL2* genes.

Reactive oxygen species (ROS) are out of equilibrium in ARHI, and the inner ear's metabolism gradually deteriorates. Inflammatory reactions are brought on by mtDNA release into the cytosol, which is brought on by oxidative stress and mtDNA damage. ARHI mice had higher levels of inflammatory cytokines in their inner ears¹⁰. To identify the vital role of apoptosis in the cochlea, many researchers usually use naturally aged C57BL/6J mice¹¹. Exploring the intracellular mechanism of cochlear ageing is crucial to understanding cell death in ARHI, and the methods that can remedy this impairment hold equal weight.

The present study aims to unravel the molecular pathways that underlie ARHI and pinpoint possible biomarkers for early diagnosis and therapeutic intervention. To achieve this, we examine the expression of *BAK1* and *BCL2* in blood samples from Pakistani patients with ARHI compared to healthy individuals, aiming to elucidate the role of apoptosis in the progression of ARHI and identify potential molecular targets for therapeutic intervention.

METHODOLOGY

This case-controlled study was conducted between 2020 and 2021 at the Department of Bioinformatics & Biotechnology, Government College Universitv Faisalabad. Sixty ARHI patients and 40 healthy subjects were selected for this study. The affected individuals were 66±3, and the healthy subjects were 30±5 years old. All the subjects were recruited from two cities in Pakistan, Faisalabad and Chichawatni. Before enrolling for this study, the participants signed the informed consent forms. The Ethical Review Committee of Government College University Faisalabad approved this research. All participants completed a questionnaire to ascertain the environmental factors and medical history that may have influenced their hearing abilities. Participants exposed to factors that can affect hearing were excluded from the study. These factors were any head injury or trauma, noise exposure, ototoxic medication, exposure to chemicals, diabetes, liver cirrhosis, neurological disease, cardiovascular disease, renal failure, psychiatric disease, cognitive dysfunction, middle ear disease, vestibular schwannoma or brain tumour, cancer, early-onset hearing loss, exposure to

gunfire, tinnitus, history of stroke, tobacco smoker, and alcohol users. Individuals struggling with cold or flu were also eliminated.

Audiometric Assessment

Pure tone audiometry was performed for all affected and healthy individuals. Air conduction hearing thresholds were measured at 250, 500, 1,000, 2,000, 4,000, and 8,000 Hz. Audiometric profiling revealed that all the affected individuals had moderately severe to severe hearing loss, whereas the healthy subjects had normal hearing (**Figure I**).

RNA Extraction and Quantification

One millilitre of blood was collected from all the individuals under study. After collection, blood was mixed with two millilitres of RBC lysis buffer, vortexed and centrifuged for 4 mins at 10,000 rpm. This step was repeated until a pallet of white blood cells was obtained. Later, 1 ml of Trizol was placed in each tube, vortexed, and kept at room temperature for five minutes. After adding the 50 µl chloroform to this lysate, the tube was kept at room temperature for 15 seconds and centrifugation at 12,000 rpm for 10 minutes. The transparent supernatant was transferred to a fresh 1.5ml tube and centrifuged at 12,000 rpm for 15 mins after adding the 500 µl isopropanol. The supernatant was removed, and the RNA pellet was washed with 500 µl of 70% ethanol and diluted with 20µl of DEPC-treated water. Isolated RNA was quantified by the nanodrop method.

cDNA Synthesis

Maxima H Minus First Strand cDNA Synthesis Kit (Thermo Scientific, cat #K1652) synthesized cDNA. 1µg of RNA of each sample and 1µl of oligo (dT) primers were added in PCR tubes separately. RNAfree water was introduced to make the total volume of 12µl in each tube. The tubes were incubated for 5 minutes at 65°C in a thermocycler. After incubation, they were chilled on ice immediately, and RT reaction reagents were added. Later, tubes were incubated for 1 hour at 43°C, and the reaction was completed by heating for 5 minutes at 70°C.

Quantitative Real-Time PCR (qPCR)

Maxima SYBR Green /ROX qPCR Master Mix 2X (Thermo Scientific, Cat number K0222) was applied for qPCR, and the expression of *BAK1*, *BCL2* and *ACTB* genes was determined using the specific primers (**Table I**). For each reaction, 5 μ I of SYBR green mix, 0.5 μ I of each primer (10 μ M), 1 μ I cDNA, and 2.4 μ I of H₂O were added. Later, a BioRAD qPCR machine was used to amplify the interested genes. The conditions for the PCR cycle were as follows: 1 cycle of denaturation at 95°C for 15 min, then 40

Table I: PCR primer sequences for BAK1, BCL2 and ACTB genes

Accession no.	. Gene	Reverse primer	Forward primer	Amplicon size
NM_001101.3	ACTB	GTA TCG GTT TGG ATG CA CCA	TCC GAG CTG AAG TACG AGC	131
NM_001188.3	BAK1	CAA ACA GGC TGG CAA TC TGG	TCA TCG GGG ACG TCA AC ACA	120
NM_000633.2	BCL2	CAG GGA GAA ATC AAA AGG CCA C	CAG ATC CTG TGG ATG GAG GCC ACT	129

cycles at 95°C for 20 sec. and 60° C for 60sec before the extension at 72°C for 60sec. then Final extension for 10min at 72°C.

RESULTS

Expression profiling of BAK1 and BCL2 genes

The expression of the apoptotic genes was quantified using the qPCR. BAK1 and BCL2 expression profiles were estimated in healthy individuals and ARHI patients. The expression of these genes was relatively compared to the expression of ACTB, a housekeeping gene. A higher expression of BAK1 in the ARHI patients was observed than in healthy subjects (Figure IIa). An upregulation of about 4.2-fold was revealed in patients in contrast to normal individuals, indicating a potential impact of the pro-apoptotic gene in hearing impairment. The expression level of the anti -apoptotic gene BCL2 was also determined. A relative increase in the BCL2 gene expression was also observed in the patients compared to healthy individuals (Figure IIb). A 2.7-fold increase in the expression level of the BCL2 gene was observed in the patients compared to the healthy subjects.

A relative expression level of anti-apoptotic and proapoptotic genes is required in the body to regulate the process of apoptosis. An increase in the pro-apoptotic genes compared to the anti-apoptotic genes will lead to apoptosis. A ratio of *BAK1/BCL2* gene expression in both patients and healthy individuals indicated an increased production of *the BAK1* gene (1.6 fold) in ARHI patients compared to the normal individuals (**Figure III**).

Figure I: The mean hearing threshold of affected and healthy individuals

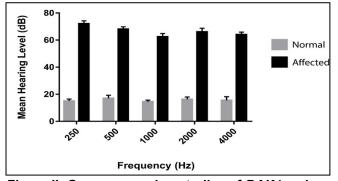
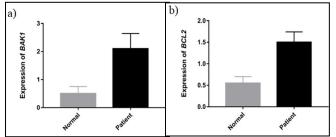
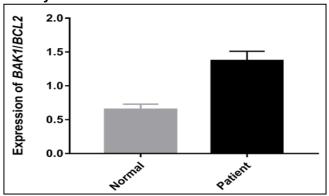


Figure II: Gene expression studies of *BAK1* and *BCL2*: a) *BAK1* gene expression in healthy and ARHI individuals. b) *BCL2* gene expression in both healthy and ARHI individuals







DISCUSSION

Presbycusis, or ARHI, is considered a progressive disorder of hearing loss in older people. This disease reduces the quality of life and has a significant societal impact¹². Age and exposure to loud noises are two factors contributing to presbycusis¹³. The burden of hearing loss is remarkably increasing among older people. Approximately two-thirds of people aged 70 and above in the United States suffer hearing loss¹⁴. The exact cause of ARHL is unknown. However, various studies have shown that oxidative stress, inflammation, apoptosis, and genetic factors are all involved in its pathophysiology. Ohlemiller KK 1999¹⁵ explored the early elevation of cochlear reactive oxygen species (ROS) and the protective effects of antioxidant treatment in acute acoustic trauma, emphasizing the role of oxidative stress in cochlear injury and potential therapeutic interventions. Hwang JH 2012¹⁶ demonstrated a relationship between reactive oxygen species (ROS) with agerelated hearing loss (ARHL). Their study revealed a notable correlation between plasma ROS levels and the extent of hearing impairment across both low and high frequencies. Additionally, Kamogashira T 2015¹⁷ further support this notion by suggesting that ROS generation contributes to apoptotic and necrotic cell death pathways in auditory tissues, which are significant factors underlying sensorineural hearing loss, including ARHL.

ARHI is controlled by various genes, which majorly contribute to the apoptosis of the inner ear cells. Upregulation of pro-apoptotic genes BAK1 and CASP3 in mice reveals hearing loss at an early age. The BAK1 gene belonged to the BCL2 protein family, which has been attributed to apoptosis. This gene family includes proapoptotic genes such as BAD and BAX and antiapoptotic genes such as BCLXL and BCL2. Members of the BCL2 protein family⁹ control permeabilization of the mitochondrial membrane during apoptosis. Apoptotic signaling in ARHL is triggered by ligands found on cell-surface receptors and takes place by mitochondria-dependent exogenous and endogenous pathways. BCL-2 protein

family members, which localize to the mitochondria, control endogenous apoptosis¹³. The BAK1 gene has a carboxyl terminus, significantly destabilizing the mitochondrial outer membrane by creating apoptotic pores. Creating the "apoptotic pore" marks the end of the mitochondrial apoptosis¹⁸. Apoptosis-related mitochondrial injury has been linked to altered morphological dynamics and mitochondrial fragmentation. Mitochondrial fragmentation involving the BAK1 gene cannot induce maximal cytochrome C release. BAK1 and BAX genes must work together to stimulate a full mitochondrial pathology. The current study demonstrated that BAK and BAX control each other during apoptosis and have unique oligomerization characteristics. During the early stages of mitochondrial outer membrane pore formation, BAK speeds up the recruitment and activation of BAX. In contrast, once inside the membrane, BAX supports the slower accumulation of more BAX molecules to expand pores. As a result, BAX and BAK co-assembly modulate mtDNA release and affect the bystander immune cells¹⁹.

Regarding BAK activation, another study reported that BH3 ligand affinity is less critical than BH3-induced changes²⁰. structural BAK1 gene-mediated mitochondrial fragmentation is decreased in BAK1 deficient mouse embryonic fibroblasts (MEF) cells²¹. In addition, the MCAT transgenic mice exhibit delayed onset of age-related HL, and investigation of the underlying molecular mechanisms revealed а decrease in the mRNA expression level of Bak1 in the cochlea. Antiapoptotic treatment has been reported to preserve hearing and reduce outer hair cell loss in mice²². A direct relationship between a higher expression of BCL2 and BCL-XL and the ageing of the inner ear mouse model has also been identified.

The most well-known model of accelerated ARHI is C57BL/6 mice, in which hearing loss begins at about six months and becomes severe at one year²³. Caloric restriction (CR), which slows the development of ARHI in C57BL/6J mice, lowers the level of apoptosis in the cochlea by reducing the level of the mitochondrial proapoptotic *BCL2* family member *BAK1*, is known to delay numerous elements of the ageing process in multiple species²⁴.

Our study showed an elevated expression of *the BAK1* gene in Pakistani ARHI patients compared to healthy subjects. Increased expression of *BAK1* (**Figure IIa**) supports the previous findings, suggesting this gene's crucial role in ARHI patients²². Furthermore, the expression of *the BCL2* gene (**Figure IIb**) was also increased in response to the high expression of the *BAK1* gene in ARHI patients when compared to normal subjects. A ratio of *BAK1/ BCL2* gene expression (**Figure III**) in both patients and healthy subjects indicated a 1.6-fold increase in ARHI patients, death of cochlear cells occurs due to increased expression of the *BAK1* gene.

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CONCLUSION

This study demonstrates a higher BAK1/BCL2 ratio and BAK1 gene expression in plasma samples of Pakistani ARHI patients. Furthermore, the increase in expression of the *BAK1/BCL2* ratio and *BAK1* gene supports the findings of previous studies. It also highlights the significance of targeting apoptotic genes such as *BAK1* for treating apoptosis-related disorders/ diseases like ARHI. This gene's expression profile can be applied as a novel biomarker for the early diagnosis of ARHI disorder.

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Conflict of Interest: The authors have no conflict of interest

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Consent to Participate: Verbal and written informed consent was taken from the study participants.

Data Sharing Statement: The corresponding author can provide the data proving the findings of this study on request. Privacy or ethical restrictions bound us from sharing the data publically.

AUTHOR CONTRIBUTION

Rauf A: Conceptualization, Writing original Draft Bhatti R: Methodology, Writing, Review and Editing Kausar S: Investigation, Methodology Aslam S: Formating & editing Nahid N: Conceptualization, Supervision Qasim M: Project Administration, editing & Review Resources

REFERENCES

- Song W, Cao H, Zhang D, Xu H, Zhang Q, Wang Z et al. Association between NR3C1 gene polymorphisms and age-related hearing impairment in Qingdao Chinese elderly. BMC Med Genomics. 2021; 14(1): 193. doi: 10.1186/s12920 -021-01044-4.
- Duan H, Song W, Wang W, Cao H, Wang B, Liu Y et al. A Genome-Wide Association Study of Age-Related Hearing Impairment in Middle- and Old-Aged Chinese Twins. BioMed Res Int. 2021; 2021: 3629624. doi: 10.1155/2021/3629624.
- 3. Lau K, Dimitriadis PA, Mitchell C, Martyn-St-James M, Hind D, Ray J. Age-related hearing loss and mild cognitive impairment: a meta-analysis and systematic review of population-based studies. J Laryngol Otol. 2022; 136(2): 103-18. doi: 10.1017/S0022215121004114. Epub 2021 Dec 13.
- 4. Curhan SG, Wang M, Eavey RD, Stampfer MJ,

Curhan GC. Adherence to Healthful Dietary Patterns Is Associated with Lower Risk of Hearing Loss in Women. J Nutr. 2018; 148(6): 944-51. doi: 10.1093/jn/nxy058.

- 5. Wang J, Puel JL. Presbycusis: An Update on Cochlear Mechanisms and Therapies. J Clin Med. 2020; 9(1): 218. doi: 10.3390/jcm9010218.
- Huang Q, Tang J. Age-related hearing loss or presbycusis. Eur Arch Otorhinolaryngol. 2010; 267(8): 1179-91. doi: 10.1007/s00405-010-1270-7. Epub 2010 May 13.
- Muderris T, Yar Sağlam AS, Unsal D, Mülazimoğlu S, Sevil E, Kayhan H. Efficiency of resveratrol in the prevention and treatment of agerelated hearing loss. Exp Ther Med. 2022; 23(1): 40. doi: 10.3892/etm.2021.10962. Epub 2021 Nov 12.
- Ahmed S, Al-Râwanduzi A. Cellular and molecular mechanisms involved in age- related hearing loss with focusing on oxidative stress. Central Asian J Med Pharmaceut Sci Innov. 2022; 2: 66-75. doi: 10.22034/CAJMPSI.2022.02.04
- 9. Falah M, Najafi M, Houshmand M, Farhadi M. Expression levels of the BAK1 and BCL2 genes highlight the role of apoptosis in age-related hearing impairment. Clin Interv Aging. 2016; 11: 1003-8. doi: 10.2147/CIA.S109110.
- Liu J, Chen H, Lin X, Yi J, Ye W, Wei F et al. Agerelated Activation of Cyclic GMP-AMP synthase-Stimulator of Interferon Genes Signaling in the Auditory System is Associated with Presbycusis in C57BL/6J Male Mice. Neuroscience. 2022; 481: 73-84. doi: 10.1016/j.neuroscience.2021.11.031. Epub 2021 Nov 27.
- 11. Liu H, Li F, Li X, Wu Q, Dai C. Rapamycin ameliorates age-related hearing loss in C57BL/6J mice by enhancing autophagy in the SGNs. Neurosci Lett. 2022; 772: 136493. doi: 10.1016/ j.neulet.2022.136493. Epub 2022 Jan 31.
- Fetoni AR, Pisani A, Rolesi R, Paciello F, Viziano A, Moleti A et al. Early Noise-Induced Hearing Loss Accelerates Presbycusis Altering Aging Processes in the Cochlea. Front Aging Neurosci. 2022; 14: 803973. doi: 10.3389/fnagi.2022.803 973.
- Kishimoto-Urata M, Urata S, Fujimoto C, Yamasoba T. Role of Oxidative Stress and Antioxidants in Acquired Inner Ear Disorders. Antioxidants(Basel). 2022; 11(8): 1469. doi: 10.3390/antiox11081469.
- 14. Lin FR, Thorpe R, Gordon-Salant S, Ferrucci L. Hearing loss prevalence and risk factors among older adults in the United States. J Gerontol A Biol

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Sci Med Sci. 2011; 66A(5): 582-90. doi: 10.1093/ gerona/glr002.

- Ohlemiller KK, Wright JS, Dugan LL. Early elevation of cochlear reactive oxygen species following noise exposure. Audiol Neurootol. 1999; 4(5): 229-36. doi: 10.1159/000013846.
- Hwang JH, Chen JC, Hsu CJ, Yang WS, Liu TC. Plasma reactive oxygen species levels are correlated with severity of age-related hearing impairment in humans. Neurobiol Aging. 2012; 33 (9): 1920-6. doi: 10.1016/j.neurobiolaging.2011. 10.012. Epub 2011 Dec 1.
- 17. Kamogashira T, Fujimoto C, Yamasoba T. Reactive oxygen species, apoptosis, and mitochondrial dysfunction in hearing loss. BioMed Res Int. 2015; 2015: 617207. doi: 10.1155/2015/ 617207. Epub 2015 Mar 22.
- Westphal D, Dewson G, Czabotar PE, Kluck RM. Molecular biology of Bax and Bak activation and action. Bioch Biophys. 2011; 1813(4): 521-31. doi: 10.1016/j.bbamcr.2010.12.019. Epub 2010 Dec 30.
- 19. Cosentino K, García-Sáez AJ. Bax and Bak Pores: Are We Closing the Circle? Trends Cell Biol. 2017; 27(4): 266-75. doi: 10.1016/j.tcb. 2016.11.004. Epub 2016 Dec 5.
- 20. Singh G, Guibao CD, Seetharaman J, Aggarwal A, Grace CR, McNamara DE et al. Structural basis of BAK activation in mitochondrial apoptosis initiation. Nature Commun. 2022; 13(1): 250. doi: 10.1038/s41467-021-27851-y.
- Brooks C, Wei Q, Feng L, Dong G, Tao Y, Mei L et al. Bak regulates mitochondrial morphology and pathology during apoptosis by interacting with mitofusins. Proc Natl Acad Sci U S A. 2007; 104 (28): 11649-54. doi: 10.1073/pnas.0703976104. Epub 2007 Jul 2.
- 22. Falah M, Najafi M, Houshmand M, Farhadi M. Expression levels of the BAK1 and BCL2 genes highlight the role of apoptosis in age-related hearing impairment. Clin Interv Aging. 2016; 11: 1003-8. doi: 10.2147/CIA.S109110.
- 23. Shin M, Pandya M, Espinosa K, Telang R, Boix J, Thorne PR et al. Istradefylline Mitigates Age-Related Hearing Loss in C57BL/6J Mice. Int J Mol Sci. 2021; 22(15): 8000. doi: 10.3390/ijms221 58000.
- Someya S, Xu J, Kondo K, Ding D, Salvi RJ, Yamasoba T et al. Age-related hearing loss in C57BL/6J mice is mediated by Bak-dependent mitochondrial apoptosis. Proc Natl Acad Sci U S A. 2009; 106(46): 19432-7. doi: 10.1073/pnas.090 8786106. Epub 2009 Nov 9.